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(only for continuation and divisional
applications under 37 CFR 1.53(b))

Docket No.: P-LJ 3430

Prior Application Info:

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Group/Art Unit: 1642

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This is a request for filing a
 X continuation divisional

application under 37 CFR 1.53(b), of pending prior application
serial no. 08/526,708, filed September 11, 1995 (list only
immediate prior application).

Title: MOLECULES THAT HOME TO A SELECTED ORGAN IN VIVO

Inventor(s) (full name of each inventor): Erkki Ruoslahti and
Renata Pasqualini

No abandonment of, or termination of proceedings, has occurred
in the above-identified prior application.

1. X An application based on the prior application as
filed and containing no new matter is enclosed,
consisting of:
 1 page application cover sheet
 59 pages of specification (includes claims and
abstract)
 0 sheets of drawing(s).
2. X 3 pages of a copy of the oath or declaration from
prior application (37 CFR 1.63(d)), U.S. serial
no. 08/526,708, filed September 11, 1995, is
enclosed.

3. ☐ A signed statement DELETING inventor(s) named in the prior application (37 CFR 1.63(d)(2) and 1.33(b)) is attached.
4. ☒ Nucleotide and/or Amino Acid Sequence Submission is enclosed:
☒ computer readable copy of sequence listing
☒ paper copy of sequence listing, pages 42 through 56
☒ Statement Under 37 C.F.R. § 1.821(f) and (g)
☐ Request to Use Computer Readable Form of Sequence Listing From Another Application
☐ Other:
5. ☒ Incorporation by Reference: The entire disclosure of the prior application, from which a copy of the oath or declaration is supplied under item no. 2 of this form, is considered as being part of the disclosure of the accompanying application and is hereby incorporated by reference herein.
6. ☐ Enter the unentered amendment previously filed on _____ under 37 CFR 1.116 in the prior application.
7. ☐ A preliminary amendment is enclosed.
8. Small entity status:
a. ☐ A small entity statement is enclosed.
b. ☐ A small entity statement was filed in prior application serial no. 08/526,708, and such status is still proper and desired.
c. ☐ is no longer claimed.
9. ☒ Amend the specification by:
☒ inserting before the first paragraph on page 1:
or
☐ deleting the paragraph on page 1 regarding related applications and inserting therefor:
- This application is a ☒ continuation of application U.S. Serial No. 08/526,708, filed September 11, 1995.
10. ☒ Cancel in this application original claim 12 of the prior application before calculating the filing fee. (At least one original independent claims must be retained for filing purposes.)

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Inventors: Ruoslahti and Pasqualini
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11. The prior application is assigned of record to La Jolla Cancer Research Foundation.
12. The power of attorney in the prior application is to Cathryn Campbell.
13. A copy of a change in power of attorney or authorization of agent as filed in the prior application (37 CFR 1.63(d)(4)) is enclosed.
14. X A return receipt postcard is enclosed.
15. An Initial Information Data Sheet is enclosed.
16. Also enclosed:

The filing fee is calculated below:

	NUMBER FILED		NUMBER EXTRA		RATE	FEE
TOTAL CLAIMS	11 - 20	=	0	x	\$9 \$18	0
INDEPENDENT CLAIMS	2 - 3	=	0	x	\$39 \$78	0
MULTIPLE DEPENDENT CLAIMS <u> </u> YES <u> X </u> NO					\$130 \$260	0
BASIC FEE					\$380 \$760	\$380
TOTAL FEE						\$380

17. A check in the amount of \$ is enclosed.
18. X The filing fee is not enclosed. Do not charge our deposit account.
19. The Commissioner is hereby authorized to charge fees under 37 CFR 1.16 and 1.17 which may be required or credit any overpayment to Deposit Account No. . A duplicate copy of this sheet is enclosed.

Inventors: Ruoslahti and Pasqualini
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Address all future communications to:

Cathryn Campbell
CAMPBELL & FLORES LLP
4370 La Jolla Village Drive, Suite 700
San Diego, California 92122
telephone: (619) 535-9001
facsimile: (619) 535-8949

Respectfully submitted,

Date: January 12, 1999

Andrea L. Gashler
Andrea L. Gashler
Registration No. 41,029
Agent for Applicant
CAMPBELL & FLORES LLP
4370 La Jolla Village Drive
Suite 700
San Diego, California 92122

___ inventor(s)

X attorney or agent of record

___ assignee of complete
interest

___ filed under Sec. 1.34(a)

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A P P L I C A T I O N

for

UNITED STATES LETTERS PATENT

on

MOLECULES THAT HOME TO A SELECTED ORGAN *IN VIVO*

by

Erkki Ruoslahti

and

Renata Pasqualini

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Attorneys

CAMPBELL AND FLORES

4370 La Jolla Village Drive, Suite 700

San Diego, California 92122

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MOLECULES THAT HOME TO A SELECTED ORGAN *IN VIVO*

This invention was made with government support under CA 42507, CA 62042 and Cancer Center Support Grant CA 30199 awarded by the National Institutes of Health.

5 The government has certain rights in this invention.

BACKGROUND OF THE INVENTION

FIELD OF THE INVENTION

The present invention relates generally to the fields of molecular medicine and drug delivery and, more
10 specifically, to molecules that home to a specific organ.

BACKGROUND INFORMATION

Although the effect of a particular pathology often is manifest throughout the body of the afflicted person, generally, the underlying pathology may affect
15 only a single organ or tissue. In many cases, drugs are the treatment of choice for a patient suffering a particular disease. It is rare, however, that a drug will target only the diseased tissue or organ. More commonly, drug treatment results in undesirable side
20 effects due, for example, to generalized toxic effects throughout the patient's body. The nausea, loss of hair and drop in blood count that occur as a result of treating of a cancer patient with chemotherapeutic agents are examples of the undesirable side effects that can
25 occur due to drug treatment.

The undesirable side effects that can occur when drugs are used to treat a disease most often are due to the inability of the drug to specifically target the diseased organ or tissue. For example, a cancer
30 chemotherapeutic agent that targets rapidly proliferating cells would be useful to kill rapidly dividing cancer

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cells. However, such an agent also kills normal
proliferating hematopoietic and epithelial cells. Thus,
the dose of such a drug that can be administered to a
patient is limited due to its toxic effect on normal
5 cells.

Efforts have been made to increase the target
specificity of various drugs. In some cases, a
particular cell type present in a diseased tissue or
organ may express a unique cell surface marker. In such
10 a case, an antibody can be raised against the unique cell
surface marker and a drug can be linked to antibody.
Upon administration of the drug/antibody complex to the
patient, the binding of the antibody to the cell surface
marker results in the delivery of a relatively high
15 concentration of the drug to the diseased tissue or
organ. Similar methods can be used where a particular
cell type in the diseased organ expresses a unique cell
surface receptor or a ligand for a particular receptor.
In these cases, the drug can be linked to the specific
20 ligand or to the receptor, respectively, thus providing a
means to deliver a relatively high concentration of the
drug to the diseased organ.

While linking a drug to a molecule that homes
to a particular cell type present in a diseased organ or
25 tissue provides significant advantages for treatment over
the use of a drug, alone, use of this method is severely
limited. In particular, very few cell type specific
antibodies have been described and it can be difficult
and time consuming to attempt to obtain an antibody that
30 targets an organ in a particular patient suffering a
pathology. Furthermore, few cell type specific surface
markers have been described. Even where such markers
have been described, the cells expressing the markers can
be distributed among various tissues or organs, thereby
35 limiting their usefulness as targets. Thus, it is

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important to identify specific target cell markers that are expressed in only one or a few tissues or organs and to identify molecules that specifically interact with such markers.

5 Various cell types can express unique markers and, therefore, provide potential targets for organ homing molecules. Endothelial cells, for example, which line the internal surfaces of blood vessels, can have distinct morphologies and biochemical markers in
10 different tissues. The blood vessels of the lymphatic system, for example, express various adhesion proteins that serve to guide lymphocyte homing. For example, endothelial cells present in lymph nodes express a cell surface marker that is a ligand for L-selectin and
15 endothelial cells in Peyer's patch venules express a ligand for the $\alpha_4\beta_7$ integrin. These ligands are involved in specific lymphocyte homing to their respective lymphoid organs. Thus, linking a drug to L-selectin or to the $\alpha_4\beta_7$ integrin may provide a means for targeting the
20 drug to diseased lymph nodes or Peyer's patches, respectively, provided that these molecules do not bind to similar ligands present in a significant number of other organs.

25 Although the homing molecules present in the blood vessels of non-lymphoid tissues have not been clearly defined, the ability of lymphocytes to return to the organ in which they were first stimulated indicates that organ-specific endothelial markers exist. Similarly, the homing or metastasis of particular types
30 of tumor cells to specific organs provides further evidence that organ-specific markers exist. However, there remains a need to identify other organ-specific cell markers and the molecules that bind to them.

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Methods are now available for producing large populations of molecules. In addition, methods are available for screening libraries of molecules to identify those of interest. For example, phage peptide display libraries can be used to express large numbers of peptides that can be screened *in vitro* with a particular target molecule or a cell of interest in order to identify peptides that specifically bind the target molecule or the cell. Screening of such phage display libraries has been used, for example, to identify ligands that specifically bind various antibodies and cell surface receptors.

Screening of a phage display library generally involves *in vitro* panning of the library using a purified target molecule. Phage that bind the target molecule can be recovered, individual phage can be cloned and the peptide expressed by a cloned phage can be determined. Such a peptide can be useful for delivery of a drug linked to the peptide to cells expressing the target molecule.

Unfortunately, very few target molecules that are expressed by only one or a few cell types have been identified. Furthermore, even where such a target molecule is known, it is uncertain whether a peptide that specifically binds the target molecule, as determined using an *in vitro* panning method, will bind to the target molecule *in vivo*. As a result, the identification of a peptide from a phage display library using an *in vitro* panning method essentially represents only a starting point for determining whether the identified peptide can be useful for an *in vivo* procedure. Thus, a need exists to develop *in vivo* methods for screening large numbers of molecules such as peptides in order to identify those that can home to one or more selected organs. The

present invention satisfies this need and provides related advantages as well.

SUMMARY OF THE INVENTION

5 The present invention provides peptides that selectively home to a selected organ. For example, the invention provides brain homing peptides having the motif, SRL (serine-arginine-lysine), such as the peptide CLSSRLDAC (SEQ ID NO: 3), or the motif VLR, such as the peptide WRCVLREGPAGGCAWFNRHRL (SEQ ID NO: 16). In
10 addition, the invention provides kidney homing peptides such as the peptide CLPVASC (SEQ ID NO: 21). An organ homing molecule of the invention is useful, for example, for targeting a moiety such as a drug to the selected organ or for identifying the presence of a target
15 molecule in a sample.

In addition, the invention provides methods of identifying a target molecule by detecting selective binding of the target molecule to a peptide of the invention. For example, a peptide that selectively homes
20 to a selected organ can be attached to a solid matrix for use in affinity chromatography. A sample of the organ can be obtained and passed over the affinity matrix under conditions that allow specific binding of the target molecule, which then can be collected and identified
25 using well known biochemical methods. The target molecule can be useful, for example, for raising an antibody specific for the target molecule.

DETAILED DESCRIPTION OF THE INVENTION

30 The present invention provides organ homing peptides, including brain homing peptides such as CNSRLHLRC (SEQ ID NO: 1), CENWWGDVC (SEQ ID NO: 2), WRCVLREGPAGGCAWFNRHRL (SEQ ID NO: 16) and others as shown

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in Table 1, and kidney homing peptides such as CLPVASC (SEQ ID NO: 21), CGAREMC (SEQ ID NO: 22) and others as shown in Table 2. Molecules that home specifically to one or a few selected organs were identified by screening a library using *in vivo* panning. The identified molecules are useful, for example, for targeting a desired moiety such as a drug, a toxin or a detectable label, which can be linked to the molecule, to the selected organ. *In vivo* panning provides a direct means for identifying molecules that specifically home to a selected organ and, therefore, provides a significant advantage over previous methods, which require that a molecule identified using an *in vitro* screening method subsequently be examined to determine whether it maintains its specificity *in vivo*.

As used herein, the term "library" means a collection of molecules. A library can contain a few or a large number of different molecules, varying from about ten molecules to several billion molecules or more. If desired, a molecule can be linked to a tag, which can facilitate recovery or identification of the molecule.

As used herein, the term "molecule" is used broadly to mean an organic chemical such as a drug; a peptide, including a variant or modified peptide or peptide-like molecules such as a peptidomimetic or peptoid; or a protein such as an antibody or a growth factor receptor or a fragment thereof such as an Fv, Fd or Fab fragment of an antibody, which contains a binding domain. For convenience, the term "peptide" is used broadly herein to mean peptides, proteins, fragments of proteins, peptoids, peptidomimetics and the like. A molecule can be a non-naturally occurring molecule, which does not occur in nature, but is produced as a result of *in vitro* methods, or can be a naturally occurring

molecule such as a protein or fragment thereof expressed from a cDNA library.

Methods for preparing libraries containing diverse populations of various types of molecules are well known in the art and commercially available (see, for example, Ecker and Crooke, Biotechnology 13:351-360 (1995), and Blondelle et al., Trends Anal. Chem. 14:83-92 (1995), and the references cited therein, each of which is incorporated herein by reference; see, also, Goodman and Ro, Peptidomimetics for Drug Design, in "Burger's Medicinal Chemistry and Drug Discovery" Vol. 1 (ed. M.E. Wolff; John Wiley & Sons 1995), pages 803-861, and Gordon et al., J. Med. Chem. 37:1385-1401 (1994), each of which is incorporated herein by reference). Where a molecule is a peptide, protein or fragment thereof, the molecule can be produced *in vitro* directly or can be expressed from a nucleic acid, which is produced *in vitro*. Methods of synthetic peptide and nucleic acid chemistry are well known in the art.

A library of molecules also can be produced, for example, by constructing a cDNA expression library from mRNA collected from a cell, tissue, organ or organism of interest. Methods for producing such libraries are well known in the art (see, for example, Sambrook et al., Molecular Cloning: A laboratory manual (Cold Spring Harbor Laboratory Press 1989), which is incorporated herein by reference). Preferably, the peptide encoded by the cDNA is expressed on the surface of a cell or a virus containing the cDNA. For example, cDNA can be cloned into a phage vector such as fuse 5 (see Example I), wherein, upon expression, the encoded peptide is expressed as a fusion protein on the surface of the phage.

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If desired, the selected organ can be processed using a method such as HPLC, which can provide a fraction enriched in molecules having a defined range of molecular weights or polar or nonpolar characteristics or the like. Conditions for HPLC will depend on the chemistry of the particular molecule and are well known to those skilled in the art. Similarly, methods for bulk removal of potentially interfering cellular materials such as DNA, RNA, proteins, lipids or carbohydrates are well known in the art as are methods for enriching a fraction containing an organic molecule using, for example, methods of selective extraction. For example, where a library comprises a population of diverse organic chemical molecules each linked to a unique oligonucleotide tag, such that the specific molecule is identified by determining the oligonucleotide sequence using PCR, genomic DNA can be removed from the sample of the collected organ in order to reduce the potential for

background PCR reactions. These and other methods can be useful for enriching the sample of the collected organ for the particular organ homing molecule, thereby removing potentially contaminating materials from the collected organ sample and increasing the sensitivity of detecting a molecule.

Evidence provided herein indicates that a sufficient number of organ homing molecules selectively home to a selected organ during *in vivo* panning such that the molecules readily can be identified. For example, various independent phage expressing the same peptide were identified in brain and in kidney (see Tables 1 and 2). Specifically, almost half of the kidney homing peptides that were sequenced had the amino acid sequence CLPVASC (SEQ ID NO: 21). Similarly, two peptides that homed to brain constituted about 40% of the sequenced brain homing peptides. Thus, a substantial fraction of the identified organ homing molecules can have the same structure. Furthermore, the peptide inserts of only a small number of isolated phage were determined. However, following various *in vivo* pannings for brain and kidney homing molecules, hundreds of thousands to millions of phage expressing organ homing peptides were recovered from the respective organs. These results indicate that some organ homing molecules will be present in substantial numbers in an organ following *in vivo* homing, thereby increasing the ease with which the molecules can be identified.

Ease of identification of an organ homing molecule depends on various factors, including the presence of potentially contaminating background cellular material. For example, where the organ homing molecule is an untagged peptide, a larger number must home to the organ in order to identify the specific peptides against the background of cellular protein. In contrast, a much

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smaller number of an untagged organic chemical homing molecule such as a drug is identifiable because such molecules normally are absent from or present in only small numbers in the body. In such a case, a highly sensitive method such as mass spectrometry can be used to identify an organ homing molecule. The skilled artisan will recognize that the method of identifying a molecule will depend, in part, on the chemistry of the particular molecule.

10 The molecules of a library also can be present as a conjugate, which can facilitate recovery or identification of the molecule. As used herein, the term "conjugate" or "molecule conjugate" means a molecule of the library linked to a physical, chemical or biological tag such as a plastic microbead, an oligonucleotide or a bacteriophage, respectively. The tag can provide a means to identify or recover an organ homing molecule of the invention following *in vivo* panning. For example, a conjugate can be a molecule such as a peptide linked to a unique oligonucleotide tag (see, for example, Brenner and Lerner, Proc. Natl. Acad. Sci., USA 89:5381-5383 (1992), which is incorporated herein by reference). Upon homing to an organ, the particular peptide can be identified by performing PCR on a sample of the organ containing the conjugate under conditions that allow amplification of the oligonucleotide tag (see, for example, Erlich, PCR Technology: Principles and Applications for DNA Amplification (Stockton Press 1989), which is incorporated herein by reference). By determining the oligonucleotide sequence, the identity of the peptide can be determined.

 In addition, a tag can be a support. As used herein, the term "support" means a tag having a defined surface to which a molecule can be attached. For example, a support can be a biological tag such as a

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virus or virus-like particle such as a bacteriophage ("phage"); a bacterium such as *E. coli*; or a eukaryotic cell such as a yeast, insect or mammalian cell; or can be a physical tag such as a liposome or a microbead, which
5 can be composed of a plastic, agarose, gelatin or other material. In general, a support should have a diameter less than about 10 μm to about 50 μm in its shortest dimension, such that the support can pass relatively unhindered through the capillary beds present in the
10 subject and not occlude circulation. In addition, a support can be nontoxic and biodegradable, particularly where the subject used for *in vivo* panning is not sacrificed to collect a selected organ.

Where a molecule is linked to a support, the
15 conjugate comprises the molecule attached to the surface of the support, such that the part of the molecule suspected of being able to interact with a target in a cell in the subject is positioned so as be able to participate in the interaction. For example, where the
20 molecule is suspected of being a β adrenergic agonist, the binding portion of the molecule attached to a support is positioned so it can interact with a β adrenergic receptor on a cell in the selected organ. Similarly, where the molecule is suspected of being a ligand for a
25 growth factor receptor, the molecule is positioned on the support so that it can bind the receptor.

As exemplified herein, a peptide suspected of being able to home to a selected organ such as brain was expressed as the N-terminus of a fusion protein, wherein
30 the C-terminus consisted of a phage coat protein. Upon expression of the fusion protein, the C-terminal coat protein linked the fusion protein to the surface of a phage such that the N-terminal peptide was in a position to interact with a target molecule in an organ. Thus, a
35 conjugate was formed by the linking of a peptide molecule

to a phage, wherein the phage provided a biological support and the peptide molecule is linked as a fusion protein.

As used herein, the term "*in vivo* panning" means a method of screening a library by administering the library to a subject and identifying a molecule that selectively homes to one or a few selected organs. The term "administering to a subject", when used in referring to a library of molecules or a portion thereof, is used in its broadest sense to mean that the library is delivered to a selected organ. For example, a library can be administered to a subject by injecting the library into the circulation of the subject such that the molecules can pass through the selected organ; after an appropriate period of time, circulation is terminated by sacrificing the subject or by removing a sample of the organ (see Example I). Alternatively, a cannula can be inserted into a blood vessel in the subject, such that the library is administered by perfusion for an appropriate period of time, after which the library can be removed from the circulation through the cannula or the subject can be sacrificed or the organ can be sampled to terminate circulation. Similarly, a library can be shunted through one or a few organs by cannulation of the appropriate blood vessels in the subject. It is recognized that a library also can be administered to an isolated perfused organ. Such panning in an isolated perfused organ can be useful for identifying molecules that bind to the organ and, if desired, can be used as an initial screening of a library. For example, if a kidney homing molecule is desired, a library can be perfused through an isolated kidney, then the molecules that bound to the perfused kidney can be screened by *in vivo* panning to identify a kidney homing molecule.

The *in vivo* panning method is exemplified herein by screening a phage peptide display library in mice and identifying specific peptides that selectively home to brain or to kidney (see Examples I and II).

5 However, phage libraries that display protein receptor molecules, including, for example, an antibody or an antigen binding fragment of an antibody such as an Fv, Fd or Fab fragment; a hormone receptor such as a growth factor receptor; or a cell adhesion receptor such as an integrin
10 or a selectin also can be used to practice the invention. Variants of such molecules can be constructed using well known methods such as random, site-directed or codon based mutagenesis (see Huse, U.S. Patent No. 5,264,563, issued November 23, 1993, which is incorporated herein by
15 reference). Thus, various types of phage display libraries can be screened using the disclosed *in vivo* panning method.

Phage display technology provides a means for expressing a diverse population of random or selectively
20 randomized peptides. Various methods of phage display and methods for producing diverse populations of peptides are well known in the art. For example, Ladner et al. (U.S. Patent No. 5,223,409, issued June 29, 1993, which is incorporated herein by reference) describe methods for
25 preparing diverse populations of binding domains on the surface of a phage. In particular, Ladner et al. describe phage vectors useful for producing a phage display library, as well as methods for selecting potential binding domains and producing randomly or
30 selectively mutated binding domains.

Similarly, Smith and Scott (Meth. Enzymol. 217:228-257 (1993); see, also, Scott and Smith, Science 249: 386-390 (1990), each of which is incorporated herein by reference) describe methods of producing phage peptide
35 display libraries, including vectors and methods of

diversifying the population of peptides that are expressed (see, also, Huse, WO 91/07141 and WO 91/07149, each of which is incorporated herein by reference; see, also, Example I). Phage display technology can be particularly powerful when used, for example, with a codon based mutagenesis method, which can be used to produce random peptides or randomly or desirably biased peptides (Huse, U.S. Patent No. 5,264,563, *supra*, 1993). These or other well known methods can be used to produce a phage display library, which can be subjected to the *in vivo* panning method of the invention in order to identify a peptide that homes to one or a few selected organs.

In addition, to screening a phage display library, *in vivo* panning can be used to screen various other types of libraries, including, for example, an RNA or cDNA library or a chemical library. If desired, the organ homing molecule can be in the form of a conjugate, which can facilitate recovery of the molecule from a selected organ or identification of the molecule in the organ. For example, where the molecules of a tagged chemical library are screened, the tag can be a moiety such as biotin, which can be linked directly to the molecule or can be linked to a support containing the molecules. Biotin provides a means to recover the molecule from a selected organ using an avidin or streptavidin affinity matrix. In addition, a molecule or a support containing a molecule can be linked to a hapten such as 4-ethoxy-methylene-2-phenyl-2-oxazoline-5-one (phOx), which can be bound by an anti-phOx antibody linked to a magnetic bead as a means to recover the molecule. Methods for purifying biotin or phOx labeled conjugates are known in the art and the materials for performing these procedures are commercially available (eg., Invitrogen; La Jolla CA; and Promega Corp.; Madison WI). In the case where a phage library is screened, the

phage can be recovered using methods as disclosed in Example I.

In vivo panning provides a method for directly identifying molecules that can home to one or a few selected organs. As used herein, the term "home" or "selectively home" means that a particular molecule binds relatively specifically to a target molecule present in one or few selected organs following administration to a subject. In general, selective homing is characterized, in part, by detecting at least a 2-fold (2x) greater specific binding of the molecule to the selected organ as compared to a control organ. It should be recognized that, in some cases, a molecule can localize non-specifically to an organ. Non-specific localization can be distinguished from homing by performing competition experiments such as those described in Examples II.C. and II.D.

Selective homing can be demonstrated by determining the specificity of an organ homing molecule for the selected organ as compared to a control organ. For example, a ratio of brain:kidney homing of up to 9:1 was observed for brain homing peptides (i.e., 9x greater binding to the selected organ as compared to the control organ; see Example II).

Selective homing also can be demonstrated by showing that molecules that home to a selected organ, as identified by one round of *in vivo* panning, are enriched for organ homing molecules in a subsequent round of *in vivo* panning. For example, phage expressing peptides that selectively home to brain were isolated by *in vivo* panning, then were subjected to additional rounds of *in vivo* panning. As demonstrated in Example II, phage recovered from brain after a first round of screening

showed an 8x enrichment in homing to brain as compared to kidney following a second round of screening and a 13x enrichment following a third round of *in vivo* panning.

When a peptide that selectively homes to brain was linked
5 to a moiety, i.e., a red blood cell (RBC), the peptide/RBC complex selectively homed to brain (see Example II.D.).

Due to the conserved nature of cellular
receptors and of ligands that bind a particular receptor,
10 the skilled artisan would recognize that an organ homing molecule identified using, for example, *in vivo* panning in a mouse also would bind to the corresponding target molecule in the selected organ of a human or other species. For example, an RGD-containing peptide that can
15 specifically bind to an integrin expressed by a cell in a human subject also can bind integrins expressed in a variety of species, including integrins expressed in mammalian cells such as murine and bovine cells as well as in cells of more evolutionarily distant species such
20 as *Drosophila*. The ability of an organ homing molecule identified using *in vivo* panning in an experimental animal such as a mouse readily can be examined for the ability to bind to the corresponding organ in a human subject by demonstrating, for example, that the molecule
25 also can bind specifically *in vitro* to a sample of the selected organ obtained from a human subject. Thus, routine methods can be used to confirm that an organ homing molecule identified using *in vivo* panning in an experimental animal also can bind an organ-specific
30 target molecule in a human subject.

The steps of administering the library to the subject, collecting a selected organ and identifying the molecules that home to the selected organ, comprise a single round of *in vivo* panning. Although not required,

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one or more additional rounds of *in vivo* panning generally are performed. Where an additional round of *in vivo* panning is performed, the molecules recovered from the selected organ in the previous round are administered to a subject, which can be the same subject used in the previous round, where only a part of the selected organ was collected.

By performing a second round of *in vivo* panning, the relative binding selectivity of the molecule conjugates recovered from the first round can be determined by administering the identified molecule conjugates to a subject, collecting the selected organ and a control organ, and comparing the molecules recovered from the selected organ with those recovered from a control organ. In addition, a second round of *in vivo* panning can indicate whether the molecules identified from the initially selected organ also can home to additional organs, thus defining a family of selected organs. Additional rounds of panning can be performed as desired.

Ideally, no molecules are recovered from a control organ following a second or subsequent round of *in vivo* panning. Generally, however, a proportion of the molecules also will be present in a control organ. In this case, the ratio of molecules in the selected organ as compared to the control organ (selected:control) can be determined. As described above, for example, phage that homed to brain following a first round of *in vivo* panning demonstrated a 13x enrichment in homing to the selected organ as compared to the control organ, kidney, following two additional rounds of panning (Example II).

Additional rounds of *in vivo* panning can be used to determine whether a particular molecule homes only to the selected organ or can recognize a target on the selected organ that also is expressed in one or more other organs or is sufficiently similar to the target in the originally selected organ. Where a molecule is found to direct homing to organs in addition to the originally selected organ, the organs are considered to constitute a family of selected organs. Using the method of *in vivo* panning, molecules that home to only a single selected organ and molecules that home to a family of selected organs can be defined. Such identification is expedited by collecting various organs during subsequent rounds of *in vivo* panning.

As used herein, the term "selected organ" is used in its broadest sense to mean an organ to which a molecule selectively homes. In addition, the term "organ" is used broadly to mean organ, tissue or cell type, including a cancer cell, in which case the selected organ can be a tumor such as a primary tumor or a metastatic lesion. In general, a selected organ contains a cell that expresses a particular target molecule such as a cell surface receptor to which an organ homing molecule can bind. By performing at least two rounds of *in vivo* panning, the selectivity of homing of the molecule to the selected organ can be determined (see Example II). As discussed above, however, in some cases an organ homing molecule can selectively home to more than one selected organ, in which case the molecule is considered to be able to selectively home to a family of selected organs.

The term "control organ" is used to mean an organ other than a selected organ. A control organ is characterized by the inability of an organ homing

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identified. For example, a mouse was injected with a phage peptide display library, then, after about 1 to 4 minutes, the mouse was frozen in liquid nitrogen to terminate circulation of the phage, the selected organ
5 (brain or kidney) was collected, phage present in the selected organ were recovered and peptides that selectively homed to the selected organ were identified (see Examples I and II).

In the examples provided, the animals were
10 sacrificed to collect the selected organ. It should be recognized, however, that only a part of the selected organ need be collected to recover a support containing a molecule that homes to that organ. For example, a part of the selected organ can be collected by biopsy, such
15 that a molecule such as a peptide expressed by a phage, can be administered to the same subject a second time or more, as desired. Where the molecule that is to be administered a second time to the same subject is linked, for example, to a support, the support must be nontoxic
20 and should be biodegradable, so as not to interfere with subsequent rounds of screening.

In vitro screening of phage libraries
previously has been used to identify peptides that bind to antibodies or to cell surface receptors (Smith and
25 Scott, *supra*, 1993). For example, *in vitro* screening of phage peptide display libraries has been used to identify novel peptides that specifically bound to integrin adhesion receptors (Koivunen et al., J. Cell Biol. 124:373-380 (1994a), which is incorporated herein by
30 reference) and to the human urokinase receptor (Goodson et al., Proc. Natl. Acad. Sci., USA 91:7129-7133 (1994)). However, such *in vitro* studies provide no insight as to whether a peptide that can specifically bind to a selected receptor *in vitro* also will bind the receptor *in*

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vivo or whether the binding peptide or the receptor are unique to a specific organ in the body. Furthermore, the *in vitro* methods are performed using defined, well-characterized target molecules in an artificial system.

5 For example, Goodson et al. utilized cells expressing a recombinant urokinase receptor. Thus, the *in vitro* methods require prior knowledge of the target molecule and yield little if any information regarding *in vivo* utility. In contrast, the *in vivo* panning method
10 disclosed herein requires no prior knowledge or availability of a target molecule. Thus, *in vivo* panning provides a significant advantage over previous methods by identifying molecules that selectively home *in vivo* to a target molecule present in one or a few organs.

15 Brain and kidney were selected as target organs to identify phage expressing peptides that selectively home to these selected organs because the blood vessels in these organs have unique characteristics. For example, the blood vessels in the brain form the
20 "blood-brain barrier" and express at least one specific antigen (Schlosshauer and Herzog, J. Cell Biol. 110:1261-1274 (1990)). Using *in vivo* panning, phage expressing various peptides that selectively homed to brain or to kidney, but not to both organs, were identified (see
25 Example II). Since the *in vivo* panning was terminated about one to four minutes after intravenous administration of the phage to the mice, the majority of phage likely remained in the vasculature, indicating the displayed peptides bound to endothelial cell surface
30 markers that are expressed in an organ-specific manner (see, for example, Springer, Cell 76:301-314 (1994)). Organ-specific homing of tumor cells and of lymphocytes indicates that the vascular beds in various tissues express unique target molecules. For example, the

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ability of lymphocytes to return to the organ in which they were first stimulated (Salmi et al., Proc. Natl. Acad. Sci., USA 89:11436-11440 (1992)) and the ability of particular types of tumor cells to home or metastasize to specific organs (Fidler and Hart, Science 217:998-1003 (1982)) indicate that organ-specific targets are expressed in a subject.

The identification of brain homing and kidney homing peptides demonstrates that organ-specific target molecules exist and indicates that the disclosed method of *in vivo* panning can be used to identify molecules that selectively home to other organs. For example, tumor vasculature, which undergoes active angiogenesis and contains specific markers (Brooks et al., Cell 79:1157-1164 (1994)), is a particularly attractive target. The identification of molecules that home to tumor vasculature would provide a means for directing a therapeutic agent directly to a tumor while sparing normal tissues.

Phage peptide display libraries were constructed essentially as described Smith and Scott (*supra*, 1993; see, also, Koivunen et al., Biotechnology 13:265-270 (1995), which is incorporated herein by reference). Oligonucleotides encoding peptides having substantially random amino acid sequences were synthesized based on an "NNK" codon, wherein "N" is A, T, C or G and "K" is G or T. "NNK" encodes 32 triplets, which encode the twenty amino acids and an amber STOP codon (Scott and Smith, *supra*, 1990). At least one codon encoding cysteine also was included in each oligonucleotide so that cyclic peptides could be formed through disulfide linkages (see Example I). The oligonucleotides were inserted in frame with the sequence encoding the gene III protein (gIII) in the vector fuse 5

such that a peptide-gIII fusion protein can be expressed. Following expression, the fusion protein is expressed on the surface of the phage containing the vector (Smith and Scott, *supra*, 1993; Koivunen et al., *supra*, 1994b).

5 Remarkably, following *in vivo* panning, the phage that selectively homed to brain or to kidney displayed only a few different peptide sequences. In some cases, peptides having the same amino acid sequence were encoded by phage having different oligonucleotide
10 sequences encoding the peptide. Furthermore, a family of brain homing peptides was identified, wherein each peptide in the family contained the common amino acid motif, SRL (serine-arginine-leucine), but different flanking amino acid sequences (see Table 1; SEQ ID NOS:
15 1, 3 and 5). In addition, two different peptides displayed the motif VLR (valine-leucine-arginine; see Table 1, SEQ ID NOS: 4 and 16). These results demonstrate that it is the peptide displayed by the phage, rather than some incidental mutant property of the
20 phage, that directs homing to the selected organ.

The sequences of the brain and kidney homing motifs do not reveal any significant similarities with known ligands for endothelial cell receptors nor do they resemble any sequences listed in various data banks.
25 However, some of the brain-homing motifs share a similarity with integrin-binding sequences. For example, one brain homing peptide contained an RLD sequence, which is recognized by certain integrins (Altieri et al., J. Biol. Chem. 265:12119-12122 (1990); Koivunen et al.,
30 *supra*, 1994a), and the DXXR (SEQ ID NO: 44) motif in another peptide resembles the RGD, DGR, and NGR motifs that bind to certain integrins (Ruoslahti, J. Clin. Invest. 87:1-5 (1991); Koivunen et al., *supra*, 1994a).

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Selective homing of a molecule such as a peptide or protein to a selected organ can be due to specific recognition by the peptide of a particular cell target molecule such as a cell surface receptor present on a cell in the organ. Selectivity of homing is dependent on the particular target molecule being expressed on only one or a few different cell types, such that the molecule homes to only one or a few organs. As discussed above, brain and kidney homing peptides likely are recognizing endothelial cell surface markers in the blood vessels present in these organs. However, most different cell types, particularly cell types that are unique to an organ, can express unique target molecules. Thus, in organs such as liver, spleen or lymph node, where blood circulates through sinusoids formed by the cells specific for the organ, *in vivo* panning can be useful for identifying molecules that home to the particular organ. For example, the sinusoids in liver are formed, in part, by hepatocytes, which are unique to liver. Using the methods disclosed herein, *in vivo* panning can be used to identify molecules that selectively home to hepatocytes and specific homing can be demonstrated by performing the appropriate competition experiments.

An organ homing molecule such as a brain homing (Table 1) or kidney homing (Table 2) peptide can be used to direct a moiety to a selected organ by linking the moiety to the molecule. As used herein, the term "moiety" is used broadly to mean an agent linked to the organ homing molecule. A moiety can be, for example, a detectable label such a radiolabel; a toxin such as ricin; or a drug such as a chemotherapeutic agent. Various moieties and methods for linking a moiety to a molecule are well known in the art and commercially available (see, for example, Harlow and Lane, Antibodies:

A laboratory manual (Cold Spring Harbor Laboratory Press 1988), which is incorporated herein by reference).

Linking of a moiety to an organ homing molecule for the purpose of directing homing of the moiety to the selected organ is exemplified by the linking of a brain homing peptide to a RBC, wherein the peptide directed homing of the RBC to brain (see Example II.D.). These results indicate that an organ homing molecule of the invention can be linked to other cell types in order to direct the cell type to a selected organ. For example, a tumor homing molecule identified by *in vivo* panning can be linked to a white blood cell (WBC) such as a cytotoxic T cell or a killer cell, wherein upon administration of the tumor homing molecule/WBC complex, the molecule directs homing of the WBC to the tumor, where the WBC can exert its effector function.

An organ homing molecule also can be linked to a moiety that is detectable external to the subject in order to perform an *in vivo* diagnostic imaging study. For example, *in vivo* imaging using a detectably labeled brain homing peptide can identify a region in the brain where circulation is occluded. For such studies, a gamma ray emitting radionuclide such as indium-111 or technetium-99 can be linked to a brain homing molecule and, following administration to a subject, can be detected using a solid scintillation detector. Alternatively, a positron emitting radionuclide such as carbon-11 or a paramagnetic spin label such as carbon-13 can be linked to the molecule and, following administration to a subject, the localization of the moiety/molecule can be detected using positron emission transaxial tomography or magnetic resonance imaging, respectively.

Such *in vivo* imaging methods also can be used to identify the presence of cancer in a subject by linking an appropriate moiety to a tumor homing molecule, which can recognize a unique target expressed by the tumor cells or by the blood vessels formed by angiogenesis in the tumor. Such methods can identify a primary tumor as well as a metastatic lesion, which may not be detectable using other methods. Having identified the presence of such cancer, the tumor homing molecule can be linked to a toxin or to a chemotherapeutic agent in order to direct the moiety to the tumor. Such a method can allow selective killing of the tumor, while substantially sparing normal tissues.

When administered to a subject, the molecule/moiety complex is administered as a pharmaceutical composition containing, for example, the complex and a pharmaceutically acceptable carrier. Pharmaceutically acceptable carriers are well known in the art and include, for example, aqueous solutions such as water or physiologically buffered saline or other solvents or vehicles such as glycols, glycerol, oils such as olive oil or injectable organic esters.

A pharmaceutically acceptable carrier can contain physiologically acceptable compounds that act, for example, to stabilize or to increase the absorption of the complex. Such physiologically acceptable compounds include, for example, carbohydrates, such as glucose, sucrose or dextrans, antioxidants, such as ascorbic acid or glutathione, chelating agents, low molecular weight proteins or other stabilizers or excipients. One skilled in the art would know that the choice of a pharmaceutically acceptable carrier, including a physiologically acceptable compound, depends, for example, on the route of administration of the

composition. The pharmaceutical composition also can contain an agent such as a cancer therapeutic agent.

One skilled in the art would know that a pharmaceutical composition containing an organ homing molecule can be administered to a subject by various routes including, for example, orally or parenterally, such as intravenously. The composition can be administered by injection or by intubation. The pharmaceutical composition also can be an organ homing molecule linked to liposomes or other polymer matrices, which can have incorporated therein, for example, a drug such as a chemotherapeutic agent (Gregoriadis, Liposome Technology, Vol. 1 (CRC Press, Boca Raton, FL 1984), which is incorporated herein by reference). Liposomes, for example, which consist of phospholipids or other lipids, are nontoxic, physiologically acceptable and metabolizable carriers that are relatively simple to make and administer.

For the diagnostic or therapeutic methods disclosed herein, an effective amount of the molecule/moiety complex must be administered to the subject. As used herein, the term "effective amount" means the amount of the complex that produces the desired effect. An effective amount often will depend on the moiety linked to the organ homing molecule. Thus, a lesser amount of a radiolabeled molecule can be required for imaging as compared to the amount of a drug/molecule complex administered for therapeutic purposes. An effective amount of a particular molecule/moiety for a specific purpose can be determined using methods well known to those in the art.

The route of administration of an organ homing molecule will depend, in part, on the chemical structure of the molecule. Peptides, for example, are not

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particularly useful when administered orally because they can be degraded in the digestive tract. However, methods for chemically modifying peptides to render them less susceptible to degradation by endogenous proteases or more absorbable through the alimentary tract are well known (see, for example, Blondelle et al., *supra*, 1995; Ecker and Crooke, *supra*, 1995; Goodman and Ro, *supra*, 1995). Such methods can be performed on peptides identified by *in vivo* panning. In addition, methods for preparing libraries of peptide analogs such as peptides containing D-amino acids; peptidomimetics consisting of organic molecules that mimic the structure of peptide; or peptoids such as vinylogous peptoids, are known in the art and can be used to identify molecules that home to a selected organ and are stable for oral administration.

Organ homing molecules obtained using the methods disclosed herein also can be useful for identifying a target molecule such as a cell surface receptor or a ligand for a receptor, which is recognized by the organ homing peptide. For example, an organ homing peptide can be linked to a solid support such as a chromatography matrix. The linked peptide then can be used for affinity chromatography by passing an appropriately processed sample of a selected organ over the column in order to bind a particular target molecule. The bound target molecule can be eluted from the column and can be identified and characterized by various well known methods. In addition, an organ homing peptide can be linked to a detectable moiety such as a radionuclide, a fluorescent molecule, an enzyme or biotin and can be used, for example, to screen a sample in order to detect the target molecule during various steps of purification of the target molecule.

The present invention also provides peptides that selectively home to a selected organ. For example,

the invention provides brain homing peptides such as CNSRLHLRC (SEQ ID NO: 1), CENWWGDVC (SEQ ID NO: 2), and others as shown in Table 1. In particular, the invention provides brain homing peptides comprising the motif, SRL (serine-arginine-lysine), such as the peptide CLSSRLDAC (SEQ ID NO: 3), or the motif VLR, such as the peptide WRCVLREGPAGGCAWFNRHRL (SEQ ID NO: 16). The invention also provides kidney homing peptides such as CLPVASC (SEQ ID NO: 21), CGAREMC (SEQ ID NO: 22) and other as shown in Table 2.

Cysteine residues were included in the peptides such that cyclization of the peptides could be effected. However, in some cases one or both of the cysteine residues in a peptide can be deleted without significantly affecting the organ homing activity of a peptide of the invention. Thus, a peptide having the sequence LSSRLDA (SEQ ID NO: 19; compare SEQ ID NO: 3) also can be a brain homing peptide. Similarly, the amino acid residues N-terminal and C-terminal to the first and last cysteine residues, respectively, in a peptide such as WRCVLREGPAGGCAWFNRHRL (SEQ ID NO: 16) can be dispensable without substantially altering brain homing activity of the peptide. Thus, a peptide having the sequence VLREGPAGG (SEQ ID NO: 20) also can be useful as a brain homing peptide. Methods for determining the necessity of a cysteine residue or of amino acid residues N-terminal or C-terminal to a cysteine residue for organ homing activity of a peptide of the invention are routine and well known in the art.

An organ homing peptide of the invention is useful, for example, for targeting a desired moiety to the selected organ as discussed above. In addition, a peptide of the invention can be used to identify the presence of a target molecule in a sample. As used herein, the term "sample" is used in its broadest sense

to mean a cell, tissue, organ or portion thereof that is isolated from the body. A sample can be, for example, a histologic section or a specimen obtained by biopsy or cells that are placed in or adapted to tissue culture.

- 5 If desired, a sample can be processed, for example, by homogenization, which can be an initial step for isolating the target molecule to which an organ homing molecule binds.

10 An organ homing peptide of the invention such as a brain homing peptide can be used to identify the target molecule expressed in brain. For example, a brain homing peptide can be attached to a matrix such as a chromatography matrix to produce a peptide affinity matrix. A homogenized sample of brain can be applied to
15 the peptide-affinity matrix under conditions that allow specific binding of the brain homing peptide to the target molecule (see, for example, Deutscher, Meth. Enzymol., Guide to Protein Purification (Academic Press, Inc., ed. M.P. Deutscher, 1990), Vol. 182, which is
20 incorporated herein by reference; see, for example, pages 357-379). Unbound and non-specifically bound material can be removed and the specifically bound brain-derived target molecule can be isolated in substantially purified form.

- 25 The following examples are intended to illustrate but not limit the present invention.

EXAMPLE I

IN VIVO PANNING

- 30 This example demonstrates methods for preparing a phage library and screening the library using *in vivo* panning to identify phage expressing peptides that home to a selected organ.

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A. Preparation of phage libraries:

Phage display libraries were constructed using the fuse 5 vector as described by Koivunen et al. (*supra*, 1995; see, also, Koivunen et al. Meth. Enzymol. 245:346-369 (1994b), which is incorporated herein by reference). Six libraries encoding peptides designated CX₅C (SEQ ID NO: 36), CX₆C (SEQ ID NO: 37), CX₇C (SEQ ID NO: 38), CX₉ (SEQ ID NO: 39), X₂CX₁₄CX₂ (SEQ ID NO: 40) and X₂CX₁₈ (SEQ ID NO: 41) were prepared, where "C" indicates cysteine and "X_N" indicates the given number of individually selected amino acids. These libraries can display cyclic peptides when at least two cysteine residues are present in the peptide.

Oligonucleotides were constructed such that "C" was encoded by the codon TGT and "X_N" was encoded by NNK, where "N" is equal molar mixtures of A, C, G and T, and where "K" is equal molar mixtures of G and T. Thus, the peptide represented by CX₅C (SEQ ID NO: 36) can be represented by an oligonucleotide having the sequence TGT(NNK)₅TGT (SEQ ID NO: 42). Oligonucleotides were made double stranded by 3 cycles of PCR amplification, purified and ligated to the nucleic acid encoding the gene III protein in the fuse 5 vector such that, upon expression, the peptide is present as a fusion protein at the N-terminus of the gene III protein.

The vectors were transfected by electroporation into MC1061 cells. Bacteria were cultured for 24 hr in the presence of 20 µg/ml tetracycline, then phage were collected from the supernatant by precipitation twice using polyethylene glycol. Each library contained about 5 x 10⁹ to 5 x 10¹⁴ transducing units (TU; individual recombinant phage).

B. In vivo panning of phage:

A mixture of phage libraries containing 1×10^{14} TU was diluted in 200 μ l DMEM and injected into the tail vein of anesthetized BALB/c mice (2 month old females; Jackson Laboratories; Bar Harbor ME); Avertin (0.015 ml/g) was used as anesthetic. After 1-4 minutes, mice were snap frozen in liquid nitrogen. To recover the phage, carcasses were partially thawed at room temperature for 1 hr, organs were collected and weighed, then were ground in 1 ml DMEM-PI (DMEM containing protease inhibitors (PI); phenylmethanesulfonyl fluoride (PMSF; 1 mM), aprotinin (20 μ g/ml), leupeptin (1 μ g/ml)).

Organ samples were washed 3 times with ice cold DMEM-PI containing 1% bovine serum albumin (BSA), then directly incubated with 1 ml K91-kan bacteria for 1 hr. Ten ml NZY medium containing 0.2 μ g/ml tetracycline (NZY/tet) was added to the bacterial culture, the mixture was incubated in a 37°C shaker for 1 hr, then 200 μ l aliquots were plated in agar plates containing 40 μ g/ml tetracycline (tet/agar).

Individual colonies containing phage recovered from brain or kidney were grown for 16 hr in 5 ml NZY/tet. The bacterial cultures obtained from the individual colonies were pooled and the amplified eluates were injected into mice as described above for a second round of *in vivo* panning. A third round of panning also was performed. Phage DNA was purified from individual bacterial colonies obtained from the second and the third round of *in vivo* panning and the DNA sequences encoding the peptides expressed by selected phage were determined (see Koivunen et al., *supra*, 1994b).

EXAMPLE II

CHARACTERIZATION OF PEPTIDES THAT HOME
TO A SELECTED ORGAN

5 This example demonstrates that an organ homing peptide of the invention selectively homes to a selected organ and that an organ homing peptide identified by *in vivo* panning can be used to direct a moiety to a selected organ.

A. Brain is the selected organ

10 Three rounds of *in vivo* panning in mice were performed. Kidney was used as a control organ. Mice were injected with two different mixtures of phage libraries. The first mixture contained libraries encoding CX₉ (SEQ ID NO: 39), CX₅C (SEQ ID NO: 36), CX₆C
15 (SEQ ID NO: 37) and CX₇C (SEQ ID NO: 38) peptides (CX₅₋₇/CX₉ mixture; SEQ ID NOS: 36-39). The second mixture contained libraries encoding X₂CX₁₄CX₂ (SEQ ID NO: 40) and X₂CX₁₈ (SEQ ID NO: 41) peptides (X₂CX₁₈/X₂CX₁₄CX₂ mixture; SEQ ID NOS: 40 and 41).

20 The phage library mixtures were administered to mice via tail vein injection. Phage input was 1×10^{16} TU of the CX₅₋₇/CX₉ (SEQ ID NOS: 36-39) mixture or 1×10^{14} TU of the X₂CX₁₈/X₂CX₁₄CX₂ (SEQ ID NOS: 40 and 41) mixture. Phage were recovered from the brains of the injected
25 mice, the recovered phage were amplified *in vitro*, then a second and a third round of *in vivo* panning was performed. During the second and third rounds of panning, phage were recovered from brain and from kidney and the number of TU from each organ was compared. This
30 comparison revealed that 6x more phage from the CX₅₋₇/CX₉ (SEQ ID NOS: 36-39) mixture bound to brain than to kidney

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10 The amino acid sequences were determined for
the inserts present in 73 cloned phage that were
recovered from brain during the second and third rounds
of *in vivo* panning. Peptides containing an SRL motif
predominated (36% of the clones sequenced; see SEQ ID
15 NOS: 1, 3 and 5), followed by peptides containing the VLR
motif (20.5% of the clones; see SEQ ID NOS: 4 and 16) and
the peptide CENWWGDVC (SEQ ID NO: 2; 19% of the clones;
see Table 1). Other peptides that occurred much less
frequently, but were present more than once, included
20 CGVRLGC (SEQ ID NO: 6), CKDWGRIC (SEQ ID NO: 7), CLDWGRIC
(SEQ ID NO: 8) and CTRITESC (SEQ ID NO: 9). Nine other
sequences appeared only one time each and were not
characterized further.

The SRL tripeptide motif was present in several different sequence contexts, indicating that the nucleic acids encoding the peptides were derived from a number of independent phage. These results indicate that the selection of the peptides containing the SRL motif represents the specific binding of several independent phage displaying peptides with the SRL sequence and is not an artifact due, for example, to phage amplification. In addition, in some cases, different phage expressed peptides that had the same amino acid sequence, but were encoded by oligonucleotides having different sequences, thus confirming that homing of a particular phage to an

organ is due to the specific peptide expressed on the phage.

To determine the specificity of brain homing of the individual motifs identified, phage displaying the predominant motifs were amplified individually, diluted to the same input titer and administered to mice. Following administration, brain and kidney were removed and the number of TU of phage in each organ was determined. The enrichment ratio of phage recovered from the selected organ, brain, as compared to the control organ, kidney, revealed that phage displaying one of the four most recovered peptides, CNSRLHLRC (SEQ ID NO: 1), CENWWGDVC (SEQ ID NO: 2), WRCVLREGPAGGCAWFNRHRL (SEQ ID NO: 16), CLSSRLDAC (SEQ ID NO: 3), each selectively targeted the brain as compared to the kidney.

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PEPTIDES FROM PHAGE RECOVERED FROM BRAIN

5	A.A. SEQUENCE (SEQ ID NO:)	# PHAGE DISPLAYING SAME A.A. SEQUENCE (% PHAGE)
	<u>CX₅₋₇C/CX₉ (36-39) library:</u>	
	CNSRLHLRC (1)	16 (21.9%)
10	CENWWGDVC (2)	14 (19.2%)
	CLSSRLDAC (3)	6 (8.2%)
	CVLRGGRC (4)	5 (6.8%)
	CNSRLQLRC (5)	4 (5.5%)
	CGVRLGC (6)	3 (4.1%)
15	CKDWGRIC (7)	2 (2.8%)
	CLDWGRIC (8)	2 (2.8%)
	CTRITESC (9)	2 (2.8%)
	CETLPAC (10)	1 (1.4%)
	CRTGTLFC (11)	1 (1.4%)
20	CGRSLDAC (12)	1 (1.4%)
	CRHWFDVVC (13)	1 (1.4%)
	CANAQSHC (14)	1 (1.4%)
	CGNPSYRC (15)	1 (1.4%)
	<u>X₂CX₁₈/X₂CX₁₄CX₂ (SEQ ID NOS: 40 and 41) library:</u>	
25	WRCVLREGPAGGCAWFNRHRL (16)	10 (13.7%)
	YPCGGEAVAGVSSVRTMCSE (17)	1 (1.4%)
	LNCDYQGTPATSVSVPCTV (18)	1 (1.4%)

Specifically, the ratio of selective homing (brain:kidney) was about 8 for CNSRLHLRC (SEQ ID NO: 1) and for CLSSRLDAC (SEQ ID NO: 3), 4 for CENWWGDVC (SEQ ID NO: 2) and 9 for WRCVLREGPAGGCAWFNRHRL (SEQ ID NO: 16).

- 5 Two control phage showed low binding to both brain and kidney. These results demonstrate that *in vivo* panning can be used to screen phage display libraries in order to identify phage expressing peptides that home to a selected organ.

10 B. Kidney as the selected organ

- The same methodology used to isolate phage expressing brain homing peptides was used to isolate phage expressing peptides that home to kidney. In these experiments, brain was used as the control organ. A
 15 mixture of the CX₅C (SEQ ID NO: 36) and the CX₆C (SEQ ID NO: 37) libraries was administered as described above. Homing of phage to the kidney was obtained and an approximately 3x to 7x enrichment of phage homing to kidney was observed following a second round of *in vivo*
 20 panning.

- The amino acid sequences were determined for the inserts in 48 cloned phage that homed to kidney. The peptides expressed by these phage were represented by two predominant sequences, CLPVASC (SEQ ID NO: 21; 46% of the clones sequenced) and CGAREMC (SEQ ID NO: 22; 17% of the clones; see Table 2). In addition, the peptide CKGRSSAC (SEQ ID NO: 23) appeared three times and three other peptides were present twice each. Phage expressing a
 25 CLPVASC (SEQ ID NO: 21), CGAREMC (SEQ ID NO: 22) or
 30 CKGRSSAC (SEQ ID NO: 23) peptide each exhibited selective homing to kidney; the ratio of selective kidney:brain homing was 7 for CLPVASC (SEQ ID NO: 21), 3 for CGAREMC (SEQ ID NO: 22) and 2 for CKGRSSAC (SEQ ID NO: 23).

TABLE 2

PEPTIDES FROM PHAGE RECOVERED FROM KIDNEY

5	A.A. SEQUENCE (SEQ ID NO:)	# PHAGE DISPLAYING SAME A.A. SEQUENCE (% PHAGE)
	CLPVASC (21)	22 (45.8%)
	CGAREMC (22)	8 (16.7%)
10	CKGRSSAC (23)	3 (6.2%)
	CWARAQGC (24)	2 (4.2%)
	CLGRSSVC (25)	2 (4.2%)
	CTSPGGSC (26)	2 (4.2%)
	CMGRWRLC (27)	1 (2.1%)
15	CVGECGGC (28)	1 (2.1%)
	CVAWLNC (29)	1 (2.1%)
	CRRFQDC (30)	1 (2.1%)
	CLMGVHC (31)	1 (2.1%)
	CKLLSGVC (32)	1 (2.1%)
20	CFVGHDL C (33)	1 (2.1%)
	CRCLNVC (34)	1 (2.1%)
	CKLMGEC (35)	1 (2.1%)

These results demonstrate that the *in vivo* panning method is a generally applicable method for screening a phage library to identify phage expressing peptides that home to a selected organ. Database searches did not reveal any significant homology of the brain homing or kidney homing peptides to known ligands for endothelial cell receptors.

C. Peptide homing is specific:

In order to confirm the specificity of a peptide for directing homing to a selected organ, peptide competition experiments were performed. The cyclic peptide, CLSSRLDAC (SEQ ID NO: 3), which is one of the brain homing peptides (see Table 1) was synthesized (Immunodynamics; La Jolla CA) and purified by HPLC. The effect on the homing of phage expressing CLSSRLDAC (SEQ ID NO: 3), CENWWGDVC (SEQ ID NO: 2) or WRCVLREGPAGGCAWFNRHRL (SEQ ID NO: 16) to the brain was examined in order to determine whether co-administration of the synthetic peptide affected homing of the phage.

Phage expressing CLSSRLDAC (SEQ ID NO: 3), CENWWGDVC (SEQ ID NO: 2) or WRCVLREGPAGGCAWFNRHRL (SEQ ID NO: 16) were titrated to the same concentration and 1 x 10⁸ TU was injected into mice alone, or with 100 µg purified synthetic cyclic CLSSRLDAC (SEQ ID NO: 3) peptide. The synthetic CLSSRLDAC (SEQ ID NO: 3) peptide inhibited the homing of phage expressing CLSSRLDAC (SEQ ID NO: 3) by about 60%. This result demonstrates that the homing of the phage to brain is specifically due to the expression on the phage of the CLSSRLDAC (SEQ ID NO: 3) peptide.

The synthetic CLSSRLDAC (SEQ ID NO: 3) peptide
25 also inhibited homing of phage expressing the
WRCVLREGPAGGCAWFNRHRL (SEQ ID NO: 16) peptide by about
60%, but did not affect homing of CENWWGDVC (SEQ ID
NO: 2) phage. This result indicates that the CLSSRLDAC
(SEQ ID NO: 3) and WRCVLREGPAGGCAWFNRHRL (SEQ ID NO: 16)
30 peptides can recognize the same target molecule in brain,
whereas the CENWWGDVC (SEQ ID NO: 2) peptide recognizes a
different target.

D. The brain homing peptide CLSSRLDAC (SEQ ID NO: 3)
directs red blood cells to brain

The synthetic cyclic CLSSRLDAC (SEQ ID NO: 3)
5 peptide (1 mg) was labeled with iodine-125 using the
Bolton and Hunter reagent (Amersham; Arlington Heights
IL). Labeled peptide was purified by reverse phase HPLC
on Sep-Pak™ cartridges (Waters, Millipore Corp.; Milford
PA) and the ¹²⁵I-peptide (200 µg) was coupled to 1 ml
10 glutaraldehyde-stabilized sheep RBC (Sigma Chemical Co.;
St. Louis MO) according to the manufacturer's
instructions.

Fifty µl ¹²⁵I-peptide/RBC (200,000 cpm), alone
or with 10 mM of unlabeled CLSSRLDAC (SEQ ID NO: 3),
15 which is a brain homing peptide, or CVRLNSLAC (SEQ ID
NO: 43), which has no brain homing activity, was injected
into the tail vein. After 2 min, each mouse was perfused
through the heart with 50 ml DMEM and the brain and
kidney were removed and assayed for radioactivity.

20 Approximately twice as much of the
CLSSRLDAC/RBC complex (SEQ ID NO: 3) homed to brain than
to kidney. Coadministration of unlabeled CLSSRLDAC (SEQ
ID NO: 3) with the complex essentially completely
inhibited brain homing of the complex but had no effect
25 on complex localizing to the kidney, indicating that
localization of the complex in the kidney was
non-specific. Coadministration of unlabeled CVRLNSLAC
(SEQ ID NO: 43) had no effect on selective homing of the
complex to brain and no effect on the non-specific
30 localization of the complex in the kidney. These results
demonstrate that an organ homing peptide identified using
in vivo panning can be linked to a moiety such as a blood
cell and selectively directs homing of the linked moiety
to the selected organ.

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Although the invention has been described with reference to the disclosed examples, it should be understood that various modifications can be made without departing from the spirit of the invention. Accordingly, 5 the invention is limited only by the following claims.

[illegible]

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: Ruoslahti, Erkki
Pasqualini, Renata
- (ii) TITLE OF INVENTION: Molecules That Home to a Selected Organ
In Vivo
- (iii) NUMBER OF SEQUENCES: 46
- (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: Campbell & Flores LLP
 - (B) STREET: 4370 La Jolla Village Drive, Suite 700
 - (C) CITY: San Diego
 - (D) STATE: California
 - (E) COUNTRY: United States
 - (F) ZIP: 92122
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER:
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
- (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: US 08/526,708
 - (B) FILING DATE: 11-SEP-1995
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Campbell, Cathryn A.
 - (B) REGISTRATION NUMBER: 31,815
 - (C) REFERENCE/DOCKET NUMBER: P-LJ 3430
- (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: (619) 535-9001
 - (B) TELEFAX: (619) 535-8949

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 9 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: both
- (ii) MOLECULE TYPE: peptide

092866 "01139
662109982260

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

Cys Asn Ser Arg Leu His Leu Arg Cys
1 5

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 9 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Cys Glu Asn Trp Trp Gly Asp Val Cys
1 5

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 9 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Cys Leu Ser Ser Arg Leu Asp Ala Cys
1 5

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 8 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Cys Val Leu Arg Gly Gly Arg Cys
1 5

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

09233366 011299
6622

- (A) LENGTH: 9 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Cys Asn Ser Arg Leu Gln Leu Arg Cys
1 5

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 7 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Cys Gly Val Arg Leu Gly Cys
1 5

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 8 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

Cys Lys Asp Trp Gly Arg Ile Cys
1 5

(2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 8 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

09223366 014300

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Cys Leu Asp Trp Gly Arg Ile Cys
1 5

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 8 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Cys Thr Arg Ile Thr Glu Ser Cys
1 5

(2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 7 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Cys Glu Thr Leu Pro Ala Cys
1 5

(2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 8 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

Cys Arg Thr Gly Thr Leu Phe Cys
1 5

(2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:

09228866 011299

- (A) LENGTH: 8 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Cys Gly Arg Ser Leu Asp Ala Cys
1 5

(2) INFORMATION FOR SEQ ID NO:13:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 9 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

Cys Arg His Trp Phe Asp Val Val Cys
1 5

(2) INFORMATION FOR SEQ ID NO:14:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 8 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Cys Ala Asn Ala Gln Ser His Cys
1 5

(2) INFORMATION FOR SEQ ID NO:15:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 8 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

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66210, 99882220

Cys Gly Asn Pro Ser Tyr Arg Cys
1 5

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 21 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: both

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

Trp Arg Cys Val Leu Arg Glu Gly Pro Ala Gly Gly Cys Ala Trp Phe
1 5 10 15
Asn Arg His Arg Leu
20

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: both

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

Tyr Pro Cys Gly Gly Glu Ala Val Ala Gly Val Ser Ser Val Arg Thr
1 5 10 15
Met Cys Ser Glu
20

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: both

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

Pro Cys Thr Val
20

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 7 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

(2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 9 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

(2) INFORMATION FOR SEQ ID NO:21:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 7 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: both

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

Cys Leu Pro Val Ala Ser Cys
1 5

(2) INFORMATION FOR SEQ ID NO:22:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 7 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

Cys Gly Ala Arg Glu Met Cys
1 5

(2) INFORMATION FOR SEQ ID NO:23:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 8 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

Cys Lys Gly Arg Ser Ser Ala Cys
1 5

(2) INFORMATION FOR SEQ ID NO:24:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 8 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

Cys Trp Ala Arg Ala Gln Gly Cys
1 5

(2) INFORMATION FOR SEQ ID NO:25:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 8 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

Cys Leu Gly Arg Ser Ser Val Cys
1 5

(2) INFORMATION FOR SEQ ID NO:26:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 8 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

Cys Thr Ser Pro Gly Gly Ser Cys
1 5

(2) INFORMATION FOR SEQ ID NO:27:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 8 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

Cys Met Gly Arg Trp Arg Leu Cys
1 5

(2) INFORMATION FOR SEQ ID NO:28:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 8 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

Cys Val Gly Glu Cys Gly Gly Cys
1 5

(2) INFORMATION FOR SEQ ID NO:29:

- (i) SEQUENCE CHARACTERISTICS:

09230306 011299

- (A) LENGTH: 7 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

Cys Val Ala Trp Leu Asn Cys
1 5

(2) INFORMATION FOR SEQ ID NO:30:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 7 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

Cys Arg Arg Phe Gln Asp Cys
1 5

(2) INFORMATION FOR SEQ ID NO:31:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 7 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

Cys Leu Met Gly Val His Cys
1 5

(2) INFORMATION FOR SEQ ID NO:32:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 8 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

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Cys Lys Leu Leu Ser Gly Val Cys
1 5

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 8 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: both

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

Cys Phe Val Gly His Asp Leu Cys
1 5

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 7 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

Cys Arg Cys Leu Asn Val Cys
1 5

(2) INFORMATION FOR SEQ ID NO:35:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 7 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

Cys Lys Leu Met Gly Glu Cys
1 5

(2) INFORMATION FOR SEQ ID NO:36:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 7 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:

Cys Xaa Xaa Xaa Xaa Xaa Cys
1 5

(2) INFORMATION FOR SEQ ID NO:37:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 8 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

Cys Xaa Xaa Xaa Xaa Xaa Xaa Cys
1 5

(2) INFORMATION FOR SEQ ID NO:38:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 9 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

Cys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Cys
1 5

(2) INFORMATION FOR SEQ ID NO:39:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 10 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:

Cys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
1 5 10

(2) INFORMATION FOR SEQ ID NO:40:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:

Xaa Xaa Cys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
1 5 10 15

Xaa Cys Xaa Xaa
20

(2) INFORMATION FOR SEQ ID NO:41:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 21 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:

Xaa Xaa Cys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
1 5 10 15

Xaa Xaa Xaa Xaa Xaa
20

(2) INFORMATION FOR SEQ ID NO:42:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 21 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:

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TGTNNKNNKN NKNNKNNKTG T

21

(2) INFORMATION FOR SEQ ID NO:43:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 9 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: both

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:

Cys Val Arg Leu Asn Ser Leu Ala Cys
 1 5

(2) INFORMATION FOR SEQ ID NO:44:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 4 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:

Asp Xaa Xaa Arg
 1

(2) INFORMATION FOR SEQ ID NO:45:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 5 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(ix) FEATURE:

- (A) NAME/KEY: Peptide
- (B) LOCATION: 1
- (D) OTHER INFORMATION: /note= "Xaa is about 1 to about 10 independently selected amino acids."

(ix) FEATURE:

- (A) NAME/KEY: Peptide
- (B) LOCATION: 5
- (D) OTHER INFORMATION: /note= "Xaa is about 1 to about 10 independently selected amino acids."

09233366 011399

Xaa Ser Arg Leu Xaa
1 5

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 5 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: unknown

```
(ix) FEATURE:
      (A) NAME/KEY: Peptide
      (B) LOCATION: 1
      (D) OTHER INFORMATION: /note= "Xaa is absent or is about 1
          to 10 independently selected amino acids."
```

```
(ix) FEATURE:
      (A) NAME/KEY: Peptide
      (B) LOCATION: 5
      (D) OTHER INFORMATION: /note= "Xaa is about 1 to about 20
          independently selected amino acids."
```

Xaa Val Leu Arg Xaa
1 5

CLAIMS

1. A peptide that selectively homes to brain.

2. The peptide of claim 1, said peptide having
5 the amino acid sequence:

X₁SRLX₂ (SEQ ID NO: 45),

wherein X_1 and X_2 each is about 1 to about 10 independently selected amino acids.

3. The peptide of claim 2, said peptide having
10 the amino acid sequence CLSSRLDAC (SEQ ID NO: 3).

4. The peptide of claim 2, said peptide having an amino acid sequence selected from the group consisting of CNSRLHLRC (SEQ ID NO: 1) and CNSRLQLRC (SEQ ID NO: 5).

5. The peptide of claim 1, said peptide having
15 the amino acid sequence:

X₃VLRX₄ (SEQ ID NO: 46),

wherein X_3 is absent or is about 1 to 10 independently selected amino acids and X_4 is about 1 to about 20 independently selected amino acids.

20 6. The peptide of claim 5, said peptide having
an amino acid sequence selected from the group consisting
of CVLRGGRC (SEQ ID NO: 4) and WRCVLREGPAGGCAWFNRHRL (SEQ
ID NO: 16).

7. The peptide of claim 1, said peptide having an amino acid sequence selected from the group consisting of CENWWGDVC (SEQ ID NO: 2) and CGVRLGC (SEQ ID NO: 6).

8. The peptide of claim 1, said peptide having an amino acid sequence selected from the group consisting of CKDWGRIC (SEQ ID NO: 7); CLDWGRIC (SEQ ID NO: 8); and CTRITESC (SEQ ID NO: 9).

9. A peptide that selectively homes to kidney.

10. The peptide of claim 9, said peptide having an amino acid sequence selected from the group consisting of CLPVASC (SEQ ID NO: 21); CGAREMC (SEQ ID NO: 22); and CKGRSSAC (SEQ ID NO: 23).

11. The peptide of claim 9, said peptide having an amino acid sequence selected from the group consisting of CWARAQGC (SEQ ID NO: 24); CLGRSSVC (SEQ ID NO: 25); and CTSPGGSC (SEQ ID NO: 26).

12. A molecule that homes to a selected organ, wherein said molecule is identified by *in vivo* panning, comprising the steps of:

a. administering to a subject a library of diverse molecules;

b. collecting a sample of the selected organ; and

c. identifying a molecule that homes to said selected organ.

ABSTRACT

MOLECULES THAT HOME TO A SELECTED ORGAN IN VIVO

The present invention provides molecules that home to a selected organ. For example, the invention
5 provides peptides that selectively home to brain or to kidney.

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PATENT
Our Docket: P-LJ 3430

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of:)
Ruoslahti and Pasqualini)
Serial No.: Not yet known)
Filed: Herewith)
For: MOLECULES THAT HOME TO)
A SELECTED ORGAN *IN VIVO*)

Group Art Unit: Unknown

Examiner: Unknown

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Washington, D.C. 20231

Sir:

Stephanie Fleischman

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(SIGNATURE OF PERSON MAILING PAPER OR FEE)

STATEMENT UNDER 37 C.F.R. § 1.821(f) and (g)

I hereby state that the content of the paper and
computer readable copies of the Sequence Listing, submitted in
accordance with 37 CFR § 1.821(c) and (e), respectively, are the
same.

I hereby state that the submission, filed in accordance
with 37 C.F.R. § 1.821(g) herein does not include new matter.

Respectfully submitted,

January 12, 1999

Date

Andrea L. Gashler

Andrea L. Gashler

Registration No. 41,029

Telephone: (619) 535-9001

Facsimile: (619) 535-8949

CAMPBELL & FLORES LLP
4370 La Jolla Village Drive
Suite 700
San Diego, California 92122

092336 01399

DECLARATION AND POWER OF ATTORNEY
FOR PATENT APPLICATION

As the below-named inventors, we hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am an original, first and joint inventor of the subject matter which is claimed and for which a patent is sought on the invention entitled MOLECULES THAT HOME TO A SELECTED ORGAN IN VIVO, which was filed September 11, 1995, as U.S. Serial No. 08/526,708 and identified as Attorney Docket No. P-LJ 1802.

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims.

I acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, Sec. 1.56(a).

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

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Inventors: Ruoslahti and Pasqualini
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We hereby appoint the following attorneys to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith:

CATHRYN CAMPBELL, Registration No. 31,815; CHRISTINE BELLAS, Registration No. 34,122; SUSAN M. PERKINS, Registration No. 36,405; RICHARD J. IMBRA, Registration No. 37,643; PAUL C. STEINHARDT, Registration No. 30,806; GREGORY R. HOOK, Registration No. 38,701; DAVID SPOLTER, Registration No. 36,933; DAVID A. GAY, Registration No. 39,200; BARRY S. WILSON, Registration No. 39,431; GREGORY P. EINHORN, Registration No. 38,440; and CALVIN A. FAN, Registration No. 38,444. Direct all telephone calls to Cathryn Campbell at telephone no. (619) 535-9001.

Address all correspondence to:

CATHRYN CAMPBELL
Campbell and Flores
4370 La Jolla Village Drive
Suite 700
San Diego, California 92122

Full name of first inventor: ERKKI RUOSLAHTI

Inventor's signature: 

Date: 11/16/95

Residence: Rancho Santa Fe, California


Citizenship: United States of America

Post Office Address: P.O. Box 1054
Rancho Santa Fe, California 92067

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Inventors: Ruoslahti and Pasqualini
Serial No.: 08/526,708
Date Filed: September 11, 1995
Page 3

Full name of second inventor: RENATA RASQUALINI

Inventor's signature: 

Date: November 10, 1995

Residence: Solana Beach, California

Citizenship: Brazil

Post Office Address: 707 South Sierra Ave. #29
Solana Beach, California 92075

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